

## WEST Search History

DATE: Tuesday, July 06, 2004

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	<i>DB=USPT,EPAB,JPAB,DWPI,TDBD; PLUR=YES; OP=OR</i>		
<input type="checkbox"/>	L11	L10 and 424/450.ccls.	51
<input type="checkbox"/>	L10	phospholipid adj5 micelle	300
<input type="checkbox"/>	L9	L5 and (phospholipid)	0
<input type="checkbox"/>	L8	L5 and (micelle)	0
<input type="checkbox"/>	L7	L5 and (phospholipid adj10 micelle)	0
<input type="checkbox"/>	L6	L5 and l1	0
<input type="checkbox"/>	L5	\$cyanoaziridine	32
<input type="checkbox"/>	L4	L1 and \$cyanoaziridine	0
<input type="checkbox"/>	L3	L1 and imexon	0
<input type="checkbox"/>	L2	L1 and 424/450.ccls.	41
<input type="checkbox"/>	L1	phospholipid adj3 micelle	203

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L2: Entry 28 of 41

File: USPT

Apr 21, 1998

DOCUMENT-IDENTIFIER: US 5741515 A

TITLE: Ketoprofen liposomes

Brief Summary Text (19):

Surprisingly, it has been found that ketoprofen liposomes can be prepared extremely simply by mixing ketoprofen with phospholipids at pHs of above 6, preferably 6 to 8, and subsequent pH reduction to values below 6, preferably 4 to 6. Owing to the deprotonated carboxyl group, the sodium salt of ketoprofen has amphiphilic character and is arranged together with the phospholipids, and mixed micelles of the phospholipid employed and the ketoprofen salt are formed. The free acid still present, ketoprofen, is incorporated into these mixed micelles. On dilution of this alkaline mixed micelle solution with a suitable buffer solution, the pH of the solution is reduced to a value below 6 and the proportion of deprotonated ketoprofen is decreased. As a result the mixed micelle membrane is destabilized and spontaneous formation of liposomes occurs. In this process it is in particular crucial that organic solvents (reverse phase evaporation method) or detergents which have to be removed from the formulation (detergent removal method) or energy-expensive comminution methods (hydration method, sonication method, French press etc.) do not have to be used as in many other preparation processes for liposomes. The formulation contains both the corresponding salt of the ketoprofen and the free acid, ketoprofen. The particle sizes are in the range from 80 to 200 nm.

Brief Summary Text (33):

Ketoprofen liposome gels according to the invention are prepared e.g. by incorporating ketoprofen phospholipid mixed micelles in certain hydrogels. To do this, ketoprofen is first dissolved in 1N NaOH solution and a mixed micelle dispersion is prepared in this medium by incorporation of a phospholipid, in particular of the formula (I). Suitable phospholipids are natural and synthetic phospholipids which can contain unsaturated and saturated fatty acids (example Phospholipon.RTM. E 90 purified soybean lecithin, Lipoid.RTM. E 80 purified egg yolk lecithin, Lipoid.RTM. E 100 purified egg yolk lecithin, Lipoid.RTM. S 100 purified soybean lecithin, Lipoid.RTM. E PC purified egg yolk lecithin, Epikuron.RTM. 200 SH purified hydrogenated soybean lecithin).

Current US Original Classification (1):

424/450

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L2: Entry 23 of 41

File: USPT

Jan 12, 1999

DOCUMENT-IDENTIFIER: US 5858398 A

**\*\* See image for Certificate of Correction \*\***

TITLE: Microparticulate pharmaceutical compositions

Brief Summary Text (22):

European Pat. Appl. No. 143,949 (Nakagama et al.) discloses a conventional liposome applying hydrogenated naturally occurring phospholipids to obtain a stable form of liposome. A fatty acid, preferably oleic acid at above 10 w % but preferably below 15 w % yields the most stable form of liposome capable of holding a drug within. Under these conditions, lecithin at its optimal weight percentage will form a lamellar liposome having oleic acid in the middle. However, at higher weight percentages of oleic acid, the phospholipids will form a micelle with oleic acid in the core, not a liposome. As disclosed by Wallach, Nakagama et al. applied cholesterol and tocopherol to strengthen the physical stability of the liposomal membrane, and applied negatively charged materials to achieve a slow releasing liposomal formulation within the body. After an intravenous infusion of liposomes containing 10,000 U of urokinase, a slow and sustained release of urokinase was observed in rabbits.

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L2: Entry 32 of 41

File: USPT

Jul 25, 1995

DOCUMENT-IDENTIFIER: US 5435989 A

**\*\* See image for Certificate of Correction \*\***

TITLE: Method of targeting a specific location in a body

Brief Summary Text (17):

When phospholipid micelles are introduced into the blood stream of a patient, the micelles move to the specific locations of cancerous growth in the patient's body, which may then be identified and treated. Drugs may be included in phospholipid vesicles and such drug-bearing vesicles may then be introduced into the patient's body for targeting the tumor locations.

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L2: Entry 37 of 41

File: USPT

May 28, 1991

DOCUMENT-IDENTIFIER: US 5019369 A

TITLE: Method of targeting tumors in humans

Detailed Description Text (5):

When phospholipid micelles are introduced into the blood stream, the micelles move to the specific locations of cancerous growth in the patient's body. To enhance movement of the phospholipid vesicles to the specific locations, positively charged phospholipid vesicles may first be introduced into the patient's blood stream to block the macrophages or other phagocytic cells in the patient's body. The positively charged molecules bound to such phospholipid vesicles may be an aminomannose or aminomannitol derivative of cholesterol. Concurrently or after a suitable period of time such as approximately one (1) hour, other phospholipid vesicles may be introduced into the patient's blood stream to move to the specific locations in the body. Such phospholipid vesicles may include cholesterol and may be neutral or may be positively charged as by the inclusion of a stearylamine or aminomannose or aminomannitol derivative of cholesterol or may be negatively charged as by the inclusion of a dicetyl phosphate.

Current US Cross Reference Classification (1):

424/450

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L11: Entry 44 of 51

File: USPT

May 28, 1991

DOCUMENT-IDENTIFIER: US 5019369 A

TITLE: Method of targeting tumors in humans

Detailed Description Text (5):

When phospholipid micelles are introduced into the blood stream, the micelles move to the specific locations of cancerous growth in the patient's body. To enhance movement of the phospholipid vesicles to the specific locations, positively charged phospholipid vesicles may first be introduced into the patient's blood stream to block the macrophages or other phagocytic cells in the patient's body. The positively charged molecules bound to such phospholipid vesicles may be an aminomannose or aminomannitol derivative of cholesterol. Concurrently or after a suitable period of time such as approximately one (1) hour, other phospholipid vesicles may be introduced into the patient's blood stream to move to the specific locations in the body. Such phospholipid vesicles may include cholesterol and may be neutral or may be positively charged as by the inclusion of a stearylamine or aminomannose or aminomannitol derivative of cholesterol or may be negatively charged as by the inclusion of a dicetyl phosphate.

Current US Cross Reference Classification (1):

424/450

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L11: Entry 50 of 51

File: USPT

Dec 23, 1980

DOCUMENT-IDENTIFIER: US 4241046 A

TITLE: Method of encapsulating biologically active materials in lipid vesicles

Brief Summary Text (26):

In a broad sense, the method of the invention calls for the formation first of "inverted micelles" in an organic phase and then the removal of the organic phase. The system then spontaneously reverts to a bilayer-like structure, with a large amount of aqueous phase encapsulated in large oligolamellar vesicles. The advantage of this method is that it gives high capture efficiencies of aqueous phase and provides large, stable vesicles. Phospholipids are excellent molecules for the formation of the "inverted micelles" and then the subsequent bilayer of the vesicles. More specifically, the method of the invention is carried out as follows.

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